

A parasitological survey on the feces of *Pan paniscus* Schwartz (1929) in Semi-liberty at “Lola ya Bonobo” sanctuary (Kinshasa city, DR Congo)

Koto-te-Nyiwa Ngbolua^{1,2,*}, Amédée K. Gbatea², Colette Masengo Ashande², Ruphin D. Djolu², Michaux K. Kamienge², Caroline I. Nkoy¹, Roger B. Lompoko¹

¹ University of Kinshasa, Faculty of Science, Department of Biology, P.O. Box 190, Kinshasa, Democratic Republic of the Congo.

² University of Gbadolite, P.O. Box 111, Gbadolite, Province of Nord-Ubangi, Democratic Republic of the Congo.

*Corresponding author: Koto-te-Nyiwa Ngbolua, E-mail: jpngbolua@unikin.ac.cd

Received: March 12, 2018, Accepted: April 14, 2018, Published: April 14, 2018.

ABSTRACT:

In the Congo basin forest, there is health risk for the human populations in term of cross-species pathogen transmission due to presence of many closely related primate species that overlap in their geographic ranges. Non-human primates (NHP) serve as important reservoirs of parasites that cause diseases to man as close interactions between humans and NHP create pathways for the cross-species transmission of zoonotic diseases. This work was assessed with the aim of identifying the intestinal helminthes of *Pan paniscus*. 45 stool samples were examined at the National Veterinary Laboratory (Kinshasa city, Congo DR) from May to June 2012 using direct wet mount, concentration via sodium chloride floatation and sedimentation methods. Identification of parasitic ova was done following established protocols. Results revealed that *Ankylostoma duodenale* had the highest infestation rate (19600 eggs: 86.3%), followed respectively by *Trichuris trichiura* (2900 eggs: 12.8%) and *Strongylus sp.* (200 eggs: 0.9%). The young animals were the less infected than teenagers and adults. The susceptibility variation of host to parasites helminthes could be attributed to the differences in animal immune responses to the infections. Regular parasitological examination of both humans and Bonobos for epidemiological monitoring and the medical treatment of infected animals should be carried out to prevent cross-species pathogen transmission in this sanctuary.

Keyword: Coprology, helminthes, Bonobo, zoonosis, Congo basin

INTRODUCTION

Wild animals like non-human primates (NHP) constitute a major reservoir of a broad arrange variety of parasites and humans may be most vulnerable to diseases from them because of their phylogenetic closeness. Indeed, many of parasitic and infectious diseases are zoonotic, having shifted from wildlife populations [1]. The geographic overlap was reported as the key factor for such cross-species pathogen transmission. In Democratic Republic of the Congo (DRC), the risk of disease transmission between wild primates and from wild primates to humans is greatest because this eco-region contains many closely related primate species that overlap in their geographic ranges. Infecting many hosts in such hotspot ecosystem, parasites were involved in emergence of news diseases and resurgence of eradicated diseases which pose a serious and increasing threat to human health and welfare [2-5]. Human emerging infectious diseases of animal reservoirs origin are largely reported in the literature [6]. Direct contact between wild primates and humans was reported to be associated with zoonotic diseases because of their close evolutionary relationship and geographic proximity [5]. Several works investigated the gastrointestinal helminthes of medical relevance in NHP [7-11]. These helminthes can however be responsible of zoonotic infections thus justifying regular parasitological checking/monitoring of animals. This study was carried out with the aim of identifying the naturally occurring gastrointestinal helminthes of *Pan paniscus*.

MATERIALS AND METHODS

Study area

The work site was the sanctuary called “Lola ya Bonobo” located in the south of the scrap forest of the lake “Ma Valée”, locality of Kimwenza, commune of “Mont Ngafula” in Kinshasa city (Democratic Republic of the Congo, see: figure 1) where these endemic animals live in semi-freedom.

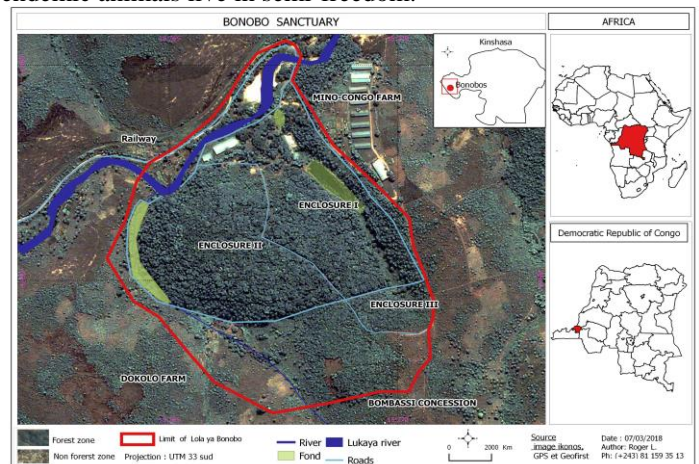


Figure 1: Geographic location of “Lola ya Bonobo” sanctuary
Collection of samples

45 feces samples were examined at the National Veterinary Laboratory (Kinshasa, Congo DR). Samples were collected between 7-9 am from May to June 2012. Top layer of fresh sample

was scooped immediately after defecation and then each sample was put in a labeled sterile bottle and kept in 10% formol. The samples were examined within 4-5 hours. The identification of the helminthes was carried out according to Halternoth & Diller [12] and Kingdon [13].

Examination of samples

Samples obtained were examined using three classical methods as previously reported [10].

Direct wet smear

Wet faecal mounts with and without staining with Lugol's iodine was used to check for the presence of parasites. One gram of the faecal sample was transferred with an applicator stick onto a grease free slide. A drop of normal saline was then added and emulsified and covers with a clean cover slip. To another slide containing one gram of the faecal sample, a drop of Lugol iodine was added and viewed under the microscope using 10x and 40x objectives. Eggs were identified based on microscopic morphology.

Simple test tube floatation

One gram of sample was put into a beaker containing 50 ml floatation fluid (40% sodium chloride) and stirred thoroughly. The

resulting suspension was filtered into labeled test tubes arranged in a rack. The test tubes were gently filled with the suspension leaving a convex meniscus on the top of the tube and a cover slip was carefully placed on top of the test tube and allowed to stand for 20 minutes. The cover slip was carefully lifted and immediately placed on a clean microscope slide and examined under the microscope at 10x and 40x objectives.

Sedimentation method

The sample was added to the normal saline solution, mixed, then washed and filtered through sieve into another beaker. The filter solution was poured into centrifuge tubes and centrifuged for 5 minutes at 1500 rpm using a centrifuge. The supernatant was decanted. One or two drop of the sediment was placed on microscope slide and viewed under a light microscope (XSP-C model) for identification of ova helminthes and adult helminthes.

Identification of helminthes

Helminthes were identified using the key books [14-17]. The microphotographs of ova were digitalized using computer assisted image analysis software (Motic Images 2000, version 1.3; Motic China Group Co LTD) [18-21].

RESULTS AND DISCUSSION

The Table 1 gives the number of parasites eggs observed.

Table 1: Number of parasites eggs observed

Age group (Feces sample)	A number of parasites eggs observed			Total	%
	<i>Ankylostoma duodenale</i>	<i>Trichuris trichiura</i>	<i>Strongylus sp</i>		
Young people (n=15)	3700	1900	200	5800	25,5
Teenager (n=15)	8200	300	00	8500	37,4
Adult (n=15)	7700	700	00	8400	37,4
Total	19600	2900	200	22700	-
%	86,3	12,8	0,9	-	100

It can deduce from this table that NHP Bonobos were infected by three species of helminthes including *Ankylostoma duodenale*, *Trichuris trichiura* and *Strongylus sp*. Adults and teenagers were equally and the most infected animals (37.4% each) followed by young animals (25.5%). *Ankylostoma duodenale* was the most abundant parasites followed respectively by *Trichuris trichiura* and *Strongylus sp*. (figure 2). Results revealed also that, young animals are specifically infected by *Strongylus sp*. The susceptibility variation of host to parasites helminthes could be attributed to the differences in animal immune responses to the infections [22].

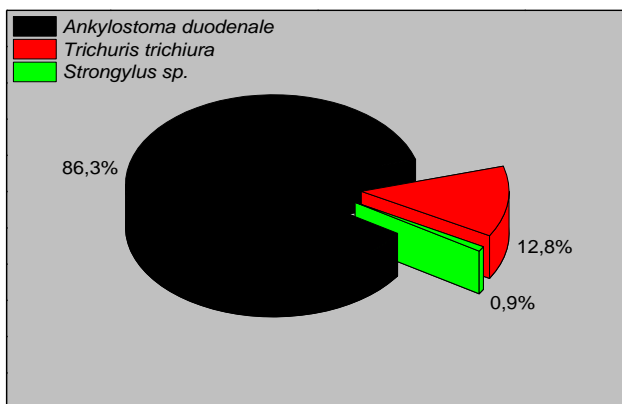


Figure 2: Occurrence of gastrointestinal helminthes in Bonobos

The figure 3 shows the image of identified helminthes eggs and the host animal *Pan paniscus*.

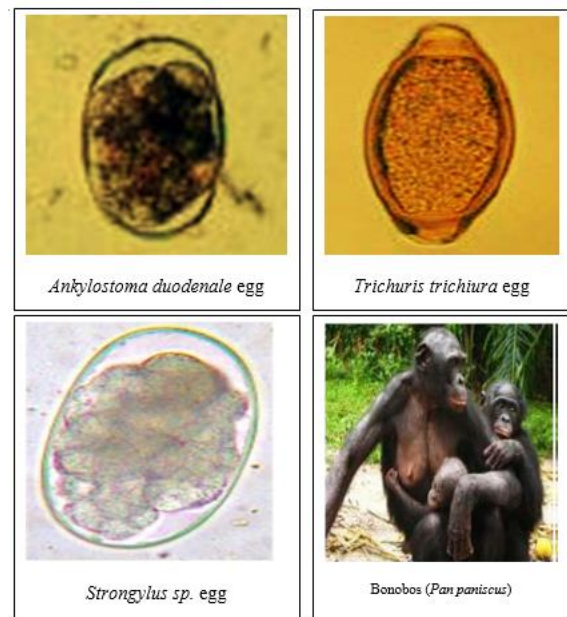


Figure 3: Image of identified helminthes eggs and the host animal *Pan paniscus*

Neglected tropical diseases (NTDs) impose a heavy socio-economic burden on the poor living in Africa by aggravating poverty and social stigma in endemic communities. The goal of current control strategies is to bring these diseases to the point where they are no longer public health problems. The main NTDs

of interest include helminthes zoonotic diseases particularly parasites-transmitted helminthiasis (PTHs). NHP were reported to be susceptible to many biological agents that infect human beings. Thus, serving as reservoirs of PTHs, NHP can transmit disease to human (zoonotic diseases) [5].

The present study provided such evidence. Indeed, in this study, NHP were found to harbor three different helminthes species of medical relevance for human (*Ankylostoma duodenale*, *Trichuris trichiura* and *Strongylus sp.*).

Among the identified helminthes, *Ankylostoma duodenale* displayed the highest infection prevalence. A similar study using other NHP like baboon and monkey as animal models revealed the occurrence of *Trichuris trichiura* (58.06%) followed by hookworm (38.71%) and *Ascaris lumbricoides* (19.35%) [10]. NHP and human, sharing the same ecosystem, there is therefore possibility of inter-specific transmission of such helminthes causing thus zoonotic diseases [9, 23].

Due to the genetic proximity between human and great ape (phylogenetic closeness), these NHP are particularly sensitive to the human illnesses. Indeed, several transmission cases of pathogenic agents from human to great ape have been reported in some eco-tourism zones [24]. The health risk also exists for the human populations particularly in the Congo basin forest in term of cross-species pathogen transmission [25]. The capacity of the NHP resilience facing the health risks in such ecosystems results in an adjustment of the self-medication behavior or zoopharmacognosy [26].

CONCLUSION

The present study revealed that Bonobos living in semi-liberty harbor zoonotic helminthes like *Ankylostoma duodenale*, *Trichuris trichiura* and *Strongylus sp.* Thus, regular parasitological examination of both humans and Bonobos for epidemiological monitoring and the medical treatment of infected animals should be carried out to prevent cross-species pathogen transmission in this sanctuary.

REFERENCES

- K.L. Vigger, D.B. Lindenmayer, D.M. Spratt, 1993. The importance of disease in reintroduction programs. *Wilfline Research* 20: 687-698.
- T.J. Davies, A.B. Pederson, 2008. Physiology and geography predict pathogen community similarity in wild primates and humans. *Proc. Boil. Sci.* 275:1695-1701.
- T.R. Gillepsie, C.A. Chapman, 2008. Forest fragmentation, the decline of an endangered primate and changes in host-parasite interactions relative to an unfragmented forest. *American Journal of Primatology* 79: 222-230.
- A. Herbert, 2009. Contribution à l'étude du parasitisme chez le mandrill au Gabon, Thèse de doctorat, Université Paul-Sabatier de Toulouse. Ecole Nationale Vétérinaire, Toulouse, France.
- A.B. Pendersen, T.J. Davies, 2010. Cross-species pathogen transmission and disease emergence in primates. *Eco-Health*. DOI: 10.1007/s10393-010-0284-3.
- K.E. Jones, N. Patel, M. Levy, A. Storeygard, D. Balk, J.L. Gittleman et al., 2008. Global trends in emerging infectious diseases. *Nature* 451: 990-993.
- E. Munene, M. Otsyul, D.A. Mbaabu, W.T. Mutahi, S.M. Muriuki, G.M. Muchemi, 1998. Helminthes and protozoan gastrointestinal tract parasites in captive and wild-trapped African Non-human Primates. *Veterinary Parasitology* 71: 73-78.
- T.R. Gillepsie, E.C. Greiner, C.A. Chapman, 2005. Gastrointestinal parasites of the Colobus monkey of Uganda, *Journal of Parasitology* 91 (3): 596-573
- B.M. Raharivololona, and J.V. Ganzhon, 2009. Gastrointestinal parasite infection of the gray mouse Lemur (*Microcebus muricus*) in the littoral forest of Mandena, Madagascar: effects of forest fragmentation and degradation. *Madagascar Conservation and development* 4 (2): 103-112.
- A. Dawet, D.P Yakubu, H. M Butu, 2013. Survey of Gastrointestinal Parasites of Non-Human Primates in Jos Zoological Garden. *Journal of Primatology* 2(1): 1-3. DOI: 10.4172/2167-6801.1000108.
- G. N. Bichitra, I. Saidul, C. Apurba, 2012. Prevalence of parasitic infection in captive non human primates of Assam State Zoo. *India Vet. World* 5 (10): 614-616.
- T.H. Halternoth, H. Diller, 1985. Mammifères d'Afrique et de Madagascar. Delacaux & Nestlé, Paris, France.
- J. Kingdon, 2009. Guide des mammifères d'Afriques. Delacaux & Nestlé, Paris, France.
- C. Chapman and M.A. Huffman, 2009. Primate and their parasites, Cambridge University Press, UK.
- C. Chartier, I. tardj, P. Morel, 2000. Précis de parasitologie vétérinaire tropicale, TEC et Doc, Editions médicales internationales, Anvers : Belgique.
- OMS, 1982. Manuel des techniques de base pour le laboratoire médical, Genève : Suisse.
- G. Lecointre and H. Guyader, 2001. Classification phylogénétique du vivant. 3éd. Bertin, Paris : France.
- Ngbolua KN, Mudogo V, Mpiana PT, Malekani MJ, Rafatro Herintsoa, Urverg Ratsimamanga U, Takoy L, Rakotoarimana H, Tshibangu DST. Evaluation de l'activité anti-drépanocytaire et antipaludique de quelques taxons végétaux de la République démocratique du Congo et de Madagascar. *Ethnopharmacologia* 2013, 50: 19-24.
- K.N. Ngbolua, T.T. Bishola, P.T. Mpiana, V. Mudogo, D.S.T. Tshibangu, K.N. Ngombe, D.D. Tshilanda, R. Baholy, 2014. In vitro antisickling and free radical scavenging activities of *Pentaclethra macrophylla* Benth. (Fabaceae). *Journal of Advancement in Medical and Life Sciences*. VII2. DOI: 10.15297/JALS.VII2.03.
- K.N. Ngbolua, T.T. Bishola, P.T. Mpiana, V. Mudogo, D.S.T. Tshibangu, K.N. Ngombe, E.G. Ekutsu, Z.B. Gbolo, N.O. Kabena, 2014. Ethno-pharmacological survey, *in vitro* antisickling and free radical scavenging activities of *Carapa procera* DC. stem bark (Meliaceae). *Nova Journal of Medical and Biological Sciences* 2(2), 01-14.
- K.N. Ngbolua, T.T. Bishola, P.T. Mpiana, V. Mudogo, D.S.T. Tshibangu, K.N. Ngombe, E.G. Ekutsu, D.D. Tshilanda, Z.B. Gbolo, D.T. Mwanangombo, P.R. Fatiany, R. Baholy, 2014. Ethno-botanical survey, *in vitro* antisickling and free radical scavenging activities of *Garcinia punctata* Oliv. (Clusiaceae). *Journal of Advanced Botany & Zoology*. VII2. DOI: 10.15297/JABZ.VII2.04.
- K.N. Ngbolua, J.M. Malekani, G.T. Tshabu., L.D. Liyandja, A.L. Pambu, L.N. Kalemba, F.B. Mwanza, D.V. Malekani, M.C. Ashande, G.N. Bongo, 2014. Epidemiological monitoring of Gastrointestinal Helminthes of two wild Galagos: *Galagoides demidovii* and *Sciurocheirus gabonensis* (Primates, Galagidae) originated from Democratic Republic of the Congo. *J. of Advancement in Medical and Life Sciences*. V2I2. DOI: 10.15297/JALS.V2I2.02.
- R. Lacost, 2009. Les parasites intestinaux chez le macaque crabier (*Macaca fuscicularis*): étude expérimentale et recommandations pour la diagnose et la gestion des rhizoflagellés et des ciliés. Thèse doctorat vétérinaire, Faculté

- de Médecine de CRETEIL, Ecole National Vétérinaire d'Alfort.
24. S. Krief, B. Vermeulen, S. Lafosse, J.M. Kasenene, A. Nieguitsila, M. Berthelemy, M. L'Hostis, O. Bain, J. Guillot, 2010. Nodular worm infection in wild chimpanzees in Western Uganda: A Risk for Human Health? PLoS Neglected Tropical Diseases 4: e630.
25. A.B. Pedersen, T.J. Davies, 2010. Cross-species pathogen transmission and disease emergence in Primates. EcoHealth 6: 496–508.
26. G.E. Ekutsu, K.N. Ngbolua, M.B. Bolaa, P.T. Mpiana, B.P. Ngoy, A.C. Masengo, G.N. Bongo, 2016. Enquête sur la pharmacopée des bonobos (*Pan paniscus*, Primates) dans un foyer endémique et Mise en évidence de l'activité anti-drépanocytaire chez un taxon végétal (*Treculia africana* Decne ex Trécul, Moraceae) testé *in vitro*. International Journal of Innovation and Applied Studies 14 (2): 315-326

Citation: Koto-te-Nyiwa Ngbolua *et al* (2018), A parasitological survey on the feces of *Pan paniscus* Schwartz (1929) in Semi-liberty at “Lola ya Bonobo” sanctuary (Kinshasa city, DR Congo). J. of Advanced Botany and Zoology. V6I2.04. DOI: 10.5281/zenodo.1217714

Copyright: © 2018 Koto-te-Nyiwa Ngbolua. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.